

**DAIRY WATERS**

**(Coliform Group and *Escherichia coli*)**  
**[*E. coli* verification required only on source water]**  
**IMS #24**

**[Unless otherwise stated all tolerances are ±5%]**

**1. Laboratory Requirements**

- a. Cultural Procedures (CP), items 33 & 34
- b. Sample volume sufficient to assure 100 mL for testing, sufficient air space for mixing (about ¾ full), if completely filled do not accept
- c. Transported and maintained at 0.0-4.5°C (temperature control [TC] required)
- d. If samples are not refrigerated, transit not to exceed 6 hours (TC not required)
- e. Transit time does not exceed 30 hours
- f. Samples examined within 30 hours of collection or within 2 hours of receipt (item 1.d)

**APPARATUS**

**2. CP, see items 1 - 32 (as necessary)**

**3. Sample Containers**

- a. Borosilicate glass, plastic bottles or bags
- b. Sterile, containing 0.1 mL of 10% Sodium Thiosulfate
- c. Holds sufficient sample with air space for all necessary bacterial tests
- d. Maintains sample uncontaminated

**4. Incubator 35±0.5°C (Make/Model: \_\_\_\_\_)**

- a. See CP item 15 for incubator requirements

**5. Water Bath, 35±0.5°C (Make/Model: \_\_\_\_\_)**

- a. Circulating and thermostatically controlled
- b. Maintain sufficient water depth

**6. Water Bath, 44.5±0.2°C (Make/Model: \_\_\_\_\_)  
[Required for EC-MUG]**

- a. Circulating and thermostatically controlled
- b. Maintain sufficient water depth

**7. Water Bath, 44.5±0.5°C (Make/Model: \_\_\_\_\_)  
[Only for use with item 31f]**

- a. Circulating and thermostatically controlled
- b. Maintain sufficient water depth

**8. Fermentation Tubes/Bottles**

- a. Sufficient size to conform with requirements for media, Durham tube and sample
- b. Tubes and bottles used for EC-MUG broth and chromogenic substrate methods do not autofluoresce

**9. Inoculation Equipment**

- a. Sterilized loops of at least 3 mm diameter, 22-24 gauge nichrome, chromel or platinum-iridium wire
- b. Disposable dry heat-sterilized hardwood applicator sticks, 0.2 to 0.3 cm in diameter and a minimum of 2.5 cm longer than the fermentation tubes
- c. Inoculating needle
- d. Sterile disposable plastic loops
- e. Commercial pre-sterilized cotton swabs on wooden sticks, 0.2 to 0.3 cm in diameter and a minimum of 2.5 cm longer than the fermentation tubes

**10. Long Wavelength UV Light (365 – 366 nm)**

- a. 6 watts
- b. Keep clean

**11. Vacuum Source with Trap**

**12. Membrane Filter (MF) Funnel; Brand: \_\_\_\_\_**

- a. Free from defects that may interfere with function
- b. Sterilizable

- c. Marked at 100 mL, or pre-marked checked and adjusted, using a 100 mL Class A graduated cylinder \_\_\_\_\_

**13. Membrane Cellulose Filters, 47 mm, 0.45 µM (±0.02 µM), Sterilized** \_\_\_\_\_

Brand: \_\_\_\_\_ Lot #: \_\_\_\_\_

**14. Absorbent Pads, Sterilized, Brand: \_\_\_\_\_** \_\_\_\_\_

**15. Forceps** \_\_\_\_\_

- a. Round tipped, with smooth surface \_\_\_\_\_

**16. Culture (Petri) Dishes (for MF), Brand: \_\_\_\_\_ Size: \_\_\_\_\_** \_\_\_\_\_

- a. Sterile with plastic, tight fitting covers \_\_\_\_\_

**17. Microscope and Lamp, Brand: \_\_\_\_\_ Model: \_\_\_\_\_** \_\_\_\_\_

- a. Binocular, wide field, 10x oculars \_\_\_\_\_
- b. Fluorescent light, adjacent, above, perpendicular to filter plane \_\_\_\_\_
- c. Other optical device giving equivalent results \_\_\_\_\_

**CULTURE MEDIA**

**18. Storage of Media** \_\_\_\_\_

- a. CP item 27 for media and storage requirements \_\_\_\_\_
- b. MF Media \_\_\_\_\_
  - 1. Store in dark at 0.0-4.5°C \_\_\_\_\_
  - 2. Broth medium used within 96 hours Date prep.: \_\_\_\_\_
  - 3. Plates kept no more than 1 week in a sealed container at 0.0-4.5°C Date prep.: \_\_\_\_\_

**19. Media Quality Control** \_\_\_\_\_

- a. See CP item 27 for media composition \_\_\_\_\_
- b. Suitability test conducted on each new lot of commercially prepared and/or each new batch of laboratory prepared media by spiking with known coliform or *E. coli*, as applicable; records maintained \_\_\_\_\_

**TESTS FOR PRESENCE OF MEMBERS OF THE COLIFORM GROUP  
AND *E. coli* BY MULTIPLE-TUBE FERMENTATION TECHNIQUE**

**20. Presumptive Test**

a. Double Strength Lauryl Sulfate Tryptose Broth (DS-LST)

1. Before inoculating arrange tubes in order and label, or otherwise identify
2. Shake samples vigorously 25 times in a 1 ft arc in 7 sec before removing test portion
3. Remove test portions (100 mL total) within 3 min
4. Inoculate ten (10) fermentation tubes containing 10 mL DS-LST or five (5) tubes containing 20 mL DS-LST or one bottle containing 100 mL DS-LST with equal volume of sample
5. Incubate tubes at  $35\pm 0.5^{\circ}\text{C}$  for  $24\pm 2$  hours
6. Examine tubes for gas - any gas is considered Presumptive Positive and must be transferred to BGLB (all tests) and EC-MUG broth (if performing *E. coli* testing)
7. Return negative tubes (no gas) to incubator and incubate an additional 24 hr (total of  $48\pm 3$  hours)
8. Re-examine tubes for gas production after  $48\pm 3$  hours
9. Record presence or absence of gas at each examination
10. Any gas produced by 24 or 48 hours is considered positive for the Presumptive Test
11. No gas after 48 hours is Not Found (NF) for the Test
  - a. Tubes showing no gas, but showing evidence of growth (turbid) can be promptly submitted to the Confirmation Test (item 21). Do not report until after completion of confirmation
12. Do not report gas production after 51 hours of incubation
13. Promptly submit all presumptive positive tubes showing gas production at 24 or 48 hours to the Confirmation Test

**21. Total Coliform Confirmation Test**

a. Brilliant Green Lactose Bile Broth (BGLB)

1. Gently shake presumptive positive tube (item 20.a.13)

2. Transfer (item 9) portion of positive broth to BGLB broth \_\_\_\_\_
3. Incubate tubes at 35±0.5°C for 24±2 hours \_\_\_\_\_
4. Examine tubes for gas - any gas is considered positive \_\_\_\_\_
5. Return negative tubes (no gas) to incubator and incubate an additional 24 hours (total of 48±3 hours) \_\_\_\_\_
6. Re-examine tubes for gas production after 48 hours \_\_\_\_\_
7. Record presence or absence of gas at each examination \_\_\_\_\_
8. Any gas produced by 24 or 48 hours is considered positive for Total Coliform \_\_\_\_\_
9. No gas after 48 hours is Not Found (NF) for Total Coliform \_\_\_\_\_
10. Do not report gas production after 51 hours of incubation \_\_\_\_\_

**22. *E. coli* Verification Test** \_\_\_\_\_

- a. EC-MUG Broth \_\_\_\_\_
  1. Gently shake presumptive positive tube(s) (item 20.a.13) \_\_\_\_\_
  2. Transfer (item 9) portion of positive broth to EC-MUG broth \_\_\_\_\_
    - a. If using the same apparatus to transfer to both BGLB and EC-MUG, transfer to EC-MUG first then to BGLB \_\_\_\_\_
  3. Incubate tubes at 44.5±0.2°C for 24±2 hours (item 6 only) \_\_\_\_\_
    - a. Place tubes in water bath within 30 min of inoculation \_\_\_\_\_
  4. Examine tubes exhibiting growth for fluorescence using Long-Wavelength UV light (item 10) \_\_\_\_\_
  5. Record presence or absence of fluorescence \_\_\_\_\_
  6. Bright blue fluorescence after incubation is considered positive for *E. coli* \_\_\_\_\_
  7. No fluorescence after incubation is Not Found (NF) for *E. coli* \_\_\_\_\_

## 23. Recording and Reporting

- a. If one or more DS-LST tubes are turbid with no gas production and confirmation in BGLB yields no gas production, invalidate the sample and request a re-sample from the same point source for heterotrophic plate count. [If history has shown a sample source to repeatedly yield growth/turbidity in LST that does not confirm, the lab may test for Heterotrophic Plate Count at the same time as testing the same sample by the Multiple Tube Fermentation technique as a usual practice.]
- b. Record results of fermentation tubes that confirm positive in BGLB as MPN Total Coliform/100 mL ( $\geq 1.1/100$  mL if 10 tubes of 10 mL or 5 tubes of 20 mL are used) or  $\geq 1$  Total Coliform/100 mL if 100 mL presence/absence test used
- c. Record results of fermentation tubes that confirm positive in EC-MUG as MPN *E. coli*/100 mL ( $\geq 1.1/100$  mL if 10 tubes of 10 mL or 5 tubes of 20 mL are used) or  $\geq 1$  *E. coli*/100 mL if 100 mL presence/absence test used
- d. Interpretation: for multiple tubes, Not Found (NF) is  $< 1.1/100$  mL and Positive is  $\geq 1.1/100$  mL; for presence/absence, NF is  $< 1/100$  mL and Positive is  $\geq 1/100$  mL

### TESTS FOR PRESENCE OF MEMBERS OF THE COLIFORM GROUP BY MEMBRANE FILTRATION TECHNIQUE

## 24. Filtration

- a. Place (with alcohol flamed forceps, item 15) sterile membrane filter (item 13) on porous plate, secure funnel
- b. Pour 100 mL test sample into funnel (item 12) and apply vacuum
- c. After test volume has been filtered, rinse funnel by filtering 3 volumes of 20-30 mL of sterile buffered water
- d. Turn off vacuum and remove filter with sterile (alcohol flamed) forceps
- e. M-Endo Broth
  - 1. Sterile pad (item 14) placed in culture dish
  - 2. Saturate pad with 2.0 mL of M-Endo Broth, CP item 27.r
  - 3. Allow to stand a few minutes before pouring off excess
  - 4. Prepared filter rolled (grid side up) onto pad slowly to avoid trapping air bubbles, do not drag across side of plate

- f. M-Endo Agar \_\_\_\_\_
  - 1. Use culture dish previously prepared (CP item 27.q) \_\_\_\_\_
  - 2. Prepared filter placed on agar with rolling motion to avoid trapping air bubbles \_\_\_\_\_

**25. Incubation** \_\_\_\_\_

- a. In saturated humidity, with dish inverted \_\_\_\_\_
- b. At  $35\pm 0.5^{\circ}\text{C}$  for  $21\pm 1$  hour \_\_\_\_\_

**26. Counting** \_\_\_\_\_

- a. Count all sheen colonies as typical coliforms and dark suspect colonies as atypical coliforms, keep separate counts of each morphological type until confirmed \_\_\_\_\_
- b. Confirm 10% up to a maximum of 10 isolated colonies, with representative proportions of each colony type \_\_\_\_\_

**27. Total Coliform Confirmation Test** \_\_\_\_\_

- a. Make serial transfers of colonies to individual LST and then to BGLB tubes using the same transfer apparatus (item 9) \_\_\_\_\_
- b. Incubate tubes at  $35\pm 0.5^{\circ}\text{C}$  for  $24\pm 2$  hours \_\_\_\_\_
- c. Examine tubes for gas \_\_\_\_\_
  - 1. LST tubes with gas must be transferred to fresh BGLB tubes if the original BGLB tubes show no gas \_\_\_\_\_
- d. Return negative tubes (no gas) to incubator and incubate an additional 24 hours (total of  $48\pm 3$  hours) \_\_\_\_\_
- e. Re-examine tubes for gas production after 48 hours \_\_\_\_\_
- f. Record presence or absence of gas at each examination \_\_\_\_\_
- g. Any gas produced in BGLB tubes by 24 or 48 hours is considered positive for the Confirmation Test \_\_\_\_\_
- h. No gas after 48 hours is Not Found (NF) for the Test \_\_\_\_\_
- i. Do not report gas production after 51 hours of incubation \_\_\_\_\_

**28. *E. coli* Verification Test**

**a. EC-MUG Broth**

- 1. Transfer (item 9) portion of each target colony to EC-MUG broth
  - a. If using the same apparatus to transfer to LTB, BGLB and EC-MUG, Transfer to LTB first, then EC-MUG then to BGLB
- 2. Incubate tubes at 44.5±0.2°C for 24±2 hours (item 6 only)
  - a. Place tube in water bath within 30 min of inoculation
- 3. Examine tubes exhibiting growth for fluorescence using Long-Wavelength UV light (item 10)
- 4. Record presence or absence of fluorescence
- 5. Bright blue fluorescence after incubation is considered positive for *E. coli*
- 6. No fluorescence after incubation is Not Found (NF) for *E. coli*

**29. Reporting**

- a. Report confirmed colony count/100 mL
- b. Invalidate all samples with confluent growth or TNTC, and request a re-sample from the same point source for heterotrophic plate count
- c. Interpretation: Not Found (NF) is < 1/100 mL and Positive is ≥ 1/100 mL

**HETEROTROPHIC BACTERIA  
STANDARD PLATE COUNT METHOD**

**30. Heterotrophic Plate Count Method**

- a. Plate samples as in SPC, items 2-10, 13 and 14
- b. Incubate at 35±0.5°C for 48±3 hours
- c. Count as in SPC item 16
- d. Report counts as in SPC item 19.a
- e. Record as "Heterotrophic Plate Count/mL at 35°C"
- f. Interpretation: Not Found (NF) if < 500 CFU/mL and Positive if ≥ 500 CFU/mL



**CHROMOGENIC SUBSTRATE (MMO-MUG) PRESENCE - ABSENCE TEST  
FOR DAIRY WATERS (SOURCE WATER SUPPLIES ONLY)**

**31. Materials**

- a. Sterile non-fluorescent borosilicate glass or clear plastic bottles to contain 100 mL sample with sufficient air space for mixing (about ¾ full)
- b. Color comparator (required for Colilert® and Colilert®-18)
- c. Commercially prepared substrate used
  - 1. Colilert® (CP item 27.s)
  - 2. Colilert®-18 (CP item 27.t) (see 32.d)
  - 3. Colisure® (CP item 27.u)
- d. Suitability test conducted on each lot of substrate received, by spiking with known coliform; maintain records
- e. Water Bath, circulating, maintains 35±0.5°C or; maintain records during periods of use (required for Colilert-18)
- f. Water Bath, circulating, maintains 44.5±0.5°C (item 6 or 7); maintain records during periods of use (optional for Colilert-18; see item 32.d)

**32. Procedure**

- a. Aseptically add pre-weighed substrate to 100 mL of the water sample
- b. Optionally, add 100 mL of sample to the substrate in a sterile container provided by the manufacturer
- c. Aseptically cap and mix thoroughly by shaking 25 times to dissolve reagent (does not completely dissolve)
- d. For Colilert-18, thermally equilibrate test solution for 20 min in a 35±0.5°C circulating water bath or alternatively 7-10 min (not to exceed 10 min) in a 44.5±0.5°C circulating water bath (item 6 or 7), and then continue incubation in water bath or dry incubator (35±0.5°C) for a total of 18 hours (minimum), not to exceed 22 hours
- e. For Colilert and Colisure, incubate at 35±0.5°C in water bath or dry incubator for a **minimum** of 24 hours, not to exceed 28 hours for Colilert, 48 hours for Colisure
- f. Examine containers for the production of color change
- g. Examine containers that exhibit color change for fluorescence

### 33. Interpretation and Reporting

a. Colilert and Colilert-18

1. If no yellow color is observed

- a. Record test result as Not Found (NF) for Total Coliform
- b. Report as Total Coliform Not Found (NF) in 100 mL sample:  
< 1/100 mL

2. If yellow color present:

- a. Gently invert container several times until color is uniformly dispersed through the sample
- b. Compare yellow color to color comparator dispersed into the **SAME** type of sample container
- c. If color is equal to or greater than that of the color comparator, record test result as Positive (POS) for Total Coliform
- d. Report as total coliform Present in 100 mL sample:  $\geq 1/100$  mL
- e. If yellow color is obvious but less than the comparator, record test result as Not Found (NF) for Total Coliform; report as for no yellow color above (33.a.1.b)

3. Place yellow containers under Long-Wavelength UV light (item 10)

- a. If the container fluoresces, record test result as Positive (POS) for *E. coli*
- b. Report as *E. coli* Present in 100 mL sample:  $\geq 1/100$  mL
- c. If container does not fluoresce, record test result as Not Found (NF) for *E. coli*
- d. Report as *E. coli* Not Found (NF) in 100 mL sample: <1/100 mL

b. Colisure

1. If no red or magenta color is observed

- a. Record test result as Not Found (NF) for Total Coliform
- b. Report as Total Coliform Not Found (NF) in 100 mL sample:  
< 1/100 mL

2. If red or magenta color present \_\_\_\_\_
  - a. Gently invert container several times until color is uniformly dispersed through the sample \_\_\_\_\_
  - b. If red or magenta color is present, record test result as Positive for Total Coliform \_\_\_\_\_
  - c. Report as Total Coliform Present in 100 mL sample:  $\geq 1/100$  mL \_\_\_\_\_
3. Place red or magenta containers under Long-Wavelength UV light \_\_\_\_\_
  - a. If the container fluoresces, record test result as Positive (POS) for *E. coli* \_\_\_\_\_
  - b. Report as *E. coli* Present in 100 mL sample:  $\geq 1/100$  mL \_\_\_\_\_
  - c. If container does not fluoresce, record test result as Not Found (NF) for *E. coli*. Report as  $< 1/100$  mL for *E. coli* \_\_\_\_\_
  - d. Report as *E. coli* Not Found (NF) in 100 mL sample:  $< 1/100$  mL \_\_\_\_\_

**CHROMOGENIC SUBSTRATE (MMO-MUG) MULTIPLE TUBE PROCEDURE  
FOR THE PRESENCE OF TOTAL COLIFORMS (SOURCE WATER SUPPLIES ONLY)**

**34. Materials** \_\_\_\_\_

- a. Sterile non-fluorescent borosilicate glass or clear plastic tubes 10 mL or 20 mL capacity \_\_\_\_\_
- b. See item 31.b (comparator solution must be in container of same size and type (34.a.)) \_\_\_\_\_
- c. See item 31.e \_\_\_\_\_

**35. Procedure** \_\_\_\_\_

- a. Before transferring sample portions arrange tubes in order and identify \_\_\_\_\_
- b. Shake sample vigorously 25 times in 7 sec with a 1 ft movement prior to adjusting to test volume \_\_\_\_\_
- c. Aseptically add pre-weighed substrate to 100 mL sample \_\_\_\_\_
- d. Optionally, add 100 mL of sample to container with substrate provided by manufacturer \_\_\_\_\_
- e. Aseptically cap and mix thoroughly by shaking 25 times to dissolve reagent (does not completely dissolve) \_\_\_\_\_
- f. Remove test portions (100 mL total) within 3 min \_\_\_\_\_

- g. Transfer 20 mL of sample/reagent mixture to five (5) tubes, or 10 mL to ten (10) tubes \_\_\_\_\_
- h. Optionally, transfer 100 mL of mixed (see item 35.b) sample to 10 tubes containing pre-dispensed substrate provided by manufacturer \_\_\_\_\_
- i. For Colilert-18, thermally equilibrate test solution for 20 min in a 35±0.5°C circulating water bath and then continue incubation in water bath or dry incubator for a total of 18 hours (minimum), not to exceed 22 hours \_\_\_\_\_
- j. For Colilert and Colisure, incubate at 35±0.5°C in water bath or dry incubator for a **minimum** of 24 hours, not to exceed 28 hours for Colilert, 48 hours for Colisure \_\_\_\_\_
- k. Examine tubes for the development of color change \_\_\_\_\_
- l. Examine tubes that exhibit color change for fluorescence \_\_\_\_\_

**36. Interpretation** \_\_\_\_\_

- a. Colilert and Colilert-18 \_\_\_\_\_
  - 1. Mix tubes to uniformly distribute yellow color \_\_\_\_\_
  - 2. Compare tubes to color comparator tube (**SAME** size and type) \_\_\_\_\_
  - 3. Record test result of tubes without color or obvious yellow color but less than comparator as Not Found (NF) for Total Coliform \_\_\_\_\_
  - 4. Record test result of tubes with yellow color equal to or greater than color comparator tube as Positive (POS) for Total Coliform \_\_\_\_\_
  - 5. Place yellow containers under Long-Wavelength UV light (item 10) \_\_\_\_\_
    - a. If the container fluoresces, record test result as Positive (POS) for *E. coli* \_\_\_\_\_
    - b. If container does not fluoresce, record test result as Not Found (NF) for *E. coli* \_\_\_\_\_
- b. Colisure \_\_\_\_\_
  - 1. Mix tubes to uniformly distribute red or magenta color \_\_\_\_\_
  - 2. Record test result of tubes without red or magenta color as Not Found (NF) for Total Coliform \_\_\_\_\_
  - 3. Record test result of tubes with red or magenta color as Positive (POS) for Total Coliform \_\_\_\_\_

4. Place red or magenta containers under Long-Wavelength UV light (item 10) \_\_\_\_\_
- a. If the container fluoresces, record test result as Positive (POS) for *E. coli* \_\_\_\_\_
  - b. If container does not fluoresce, record test result as Not Found (NF) for *E. coli* \_\_\_\_\_

**37. Reporting** \_\_\_\_\_

- a. If all tubes exhibit no color change (36.a.3 or 36.b.2), report as Not Found (NF): < 1.1/100 mL for Total Coliform and *E. coli* \_\_\_\_\_
- b. If one or more tubes exhibit color change (36.a.4 or 36.b.3), report as Positive (POS): ≥ 1.1/100 mL for Total Coliform \_\_\_\_\_
- c. If one or more tubes exhibit fluorescence, report as Positive (POS): ≥ 1.1/100 mL for *E. coli* \_\_\_\_\_

**CHROMOGENIC SUBSTRATE PRESENCE (XGAL - MUG) PRESENCE - ABSENCE TEST FOR DAIRY WATERS (SOURCE WATER SUPPLIES ONLY)**

**38. Materials** \_\_\_\_\_

- a. E\*Colite substrate, see CP item 27.v \_\_\_\_\_
- b. Quality control procedures conducted on each lot of substrate received, as recommended by manufacturer, test by spiking with known coliform, records maintained \_\_\_\_\_

**39. Procedure** \_\_\_\_\_

- a. Add water sample to the E\*Colite substrate \_\_\_\_\_
  1. Tear perforated strip \_\_\_\_\_
  2. Open bag by pulling white tabs \_\_\_\_\_
  3. Aseptically pour 100 mL of water sample into bag (do not touch inside of bag) \_\_\_\_\_
  4. Flatten bag to remove air \_\_\_\_\_
  5. Twirl bag 2-3 times around twister wires to form a leak proof seal \_\_\_\_\_
  6. Fold twistors around back of bag \_\_\_\_\_
  7. Shake bag 25 times in 7 sec to dissolve sodium thiosulfate tablet, if present \_\_\_\_\_

8. Continue rolling to build pressure in water compartment \_\_\_\_\_
  9. Maintain pressure on rolled area and push water through first seal into powder section of bag **ONLY** \_\_\_\_\_
  10. Shake bag 25 times in 7 sec to completely dissolve powder in water (push mixture against bag sides to pull apart any remaining seal) \_\_\_\_\_
- b. Place sealed bag in 35°C water bath for 10 min \_\_\_\_\_
  - c. Transfer to 35±0.5°C incubator for 28 hours \_\_\_\_\_
  - d. Examine bags for the production of blue or blue/green color or blue color in corners of bag \_\_\_\_\_

**40. Interpretation and Reporting** \_\_\_\_\_

- a. If yellow color is observed: \_\_\_\_\_
  1. Record sample as Not Found (NF) for Total Coliform \_\_\_\_\_
  2. Report as Total Coliform Not Found (NF) in 100 mL sample: < 1/100 mL \_\_\_\_\_
- b. If blue or blue/green (or blue in corners) color observed: \_\_\_\_\_
  1. The sample is Positive for Total Coliform \_\_\_\_\_
  2. Report as Total Coliform present in 100 mL sample: ≥ 1/100 mL \_\_\_\_\_
- c. Place blue or blue/green containers under Long-Wavelength UV light (item 10) \_\_\_\_\_
  1. If the container fluoresces, record test result as Positive (POS) for *E. coli* \_\_\_\_\_
  2. Report as *E. coli* Present in 100 mL sample: ≥ 1/100 mL \_\_\_\_\_
  3. If container does not fluoresce, record test result as Not Found (NF) for *E. coli*. Report as < 1/100mL for *E. coli* \_\_\_\_\_

**MISCELLANEOUS**

**41. Copy of current in-use edition of Standard Methods for the Examination of Water and Wastewater in laboratory** \_\_\_\_\_